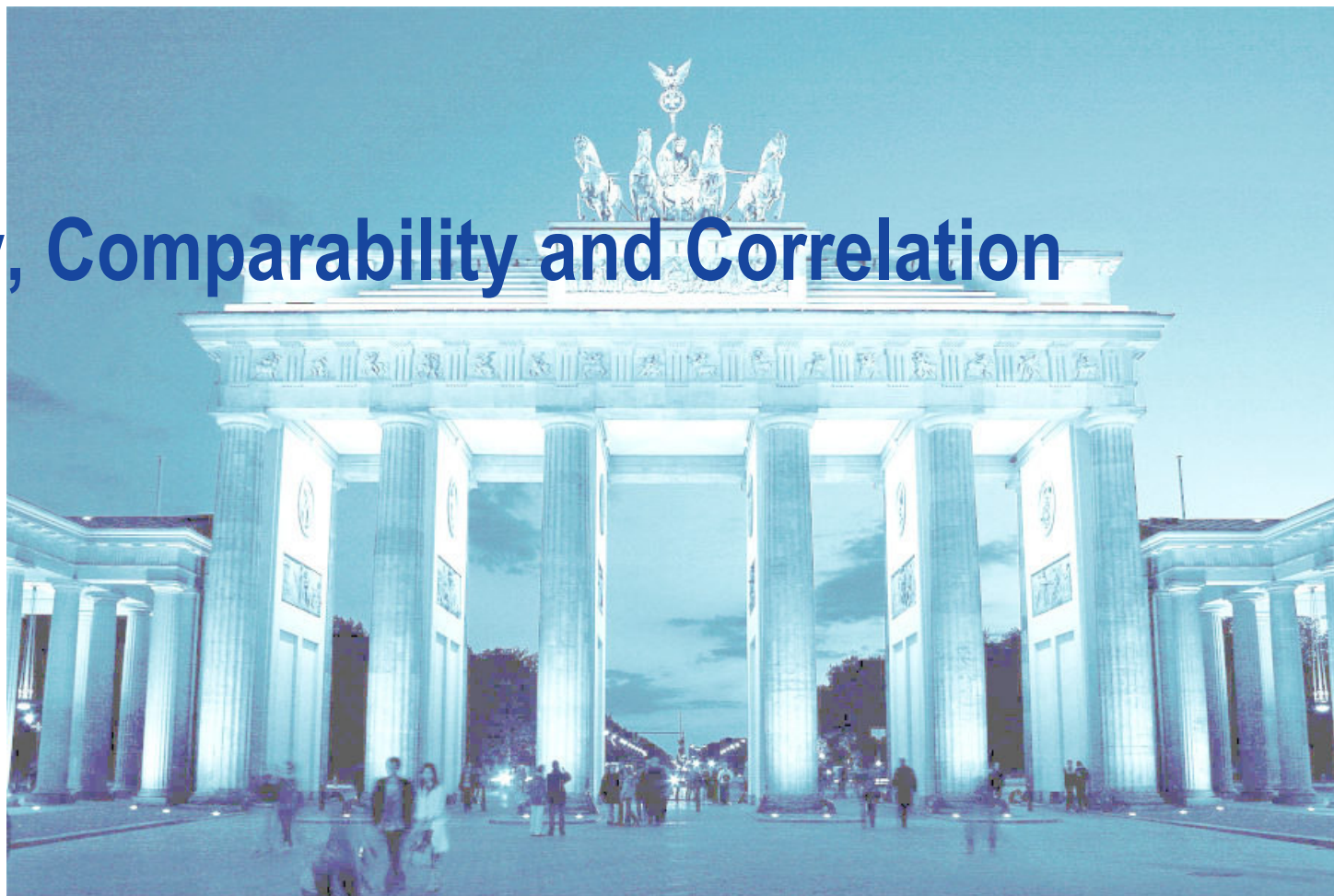


# Similarity, Comparability and Correlation

Helmut Schütz



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# Difference factor $f_1$ , similarity factor $f_2$

## Difference factor $f_1$

- Percent difference between dissolution profiles at each time point
- Measurement of the relative error between the curves.

$$f_1 = 100 \left\{ \frac{\sum_{t=1}^{t=n} |R_t - T_t|}{\sum_{t=1}^{t=n} R_t} \right\}$$

## Similarity factor $f_2$

- Logarithmic reciprocal square root transformation of the sum of squared error.
- Measurement of the similarity in the percent dissolution between the curves.

$$f_2 = 50 \cdot \log \left\{ 100 \cdot \left[ 1 / \sqrt{1 + \frac{1}{n} \sum_{t=1}^{t=n} (R_t - T_t)^2} \right] \right\}$$

# Difference factor $f_1$ , similarity factor $f_2$

## Simple example

n	3
$\Sigma (R_t - T_t)$	10
$\Sigma  R_t - T_t $	10
$\Sigma (R_t - T_t)^2$	38
$\Sigma R_t$	258
$f_2$	71.6
$f_2$	3.9

t (min)	$R_t$ (%)	$T_t$ (%)	$\Delta (R_t - T_t)$	$\Delta  R_t - T_t $	$\Delta^2$
15	83	78	5	5	25
30	85	83	2	2	4
45	90	87	3	3	9

# Difference factor $f_1$ , similarity factor $f_2$

**Certain conditions must be fulfilled for the application of  $f_2$ .**

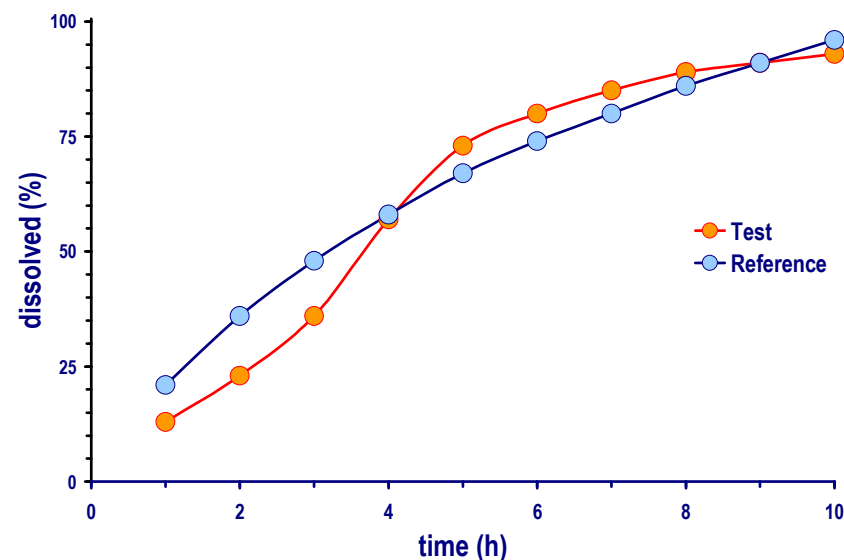
- $f_2$  *not required* if product releases  $\geq 85\%$  in all three media.
- 12 units of test and reference product.  
 $R_t$  and  $T_t$  are their arithmetic means.
- CV should not be  $>20\%$  at  $\leq 15$  minutes.
- CV should not be  $>10\%$  at other time points.
- Sampling time points after 85% release.
  - FDA: Only one measurement included for test product.
  - EMA: Not more than one mean value of  $>85\%$  dissolved for each formulation.
  - WHO: Maximum of one time-point should be considered after 85% dissolution of the comparator (Brand/Reference/Innovator) product has been reached.

# Difference factor $f_1$ , similarity factor $f_2$

## Different release characteristics

- Cave: Although  $f_1$  (2.1) and  $f_2$  (57.7) suggest similarity, the comparison is not suitable because the profiles display different release kinetics.

$t$ (h)	$R_t$ (%)	$T_t$ (%)	$\Delta (R_t - T_t)$	$\Delta  R_t - T_t $	$\Delta^2$
1	21	13	8	8	64
2	36	23	13	13	169
3	48	36	12	12	144
4	58	57	1	1	1
5	67	73	-6	6	36
6	74	80	-6	6	36
7	80	85	-5	5	25
8	86	89	-3	3	9
9	91	91			
10	96	93			



Reference: Zero order?

Test: Sigmoidal (Hill or Weibull?)

# Alternatives?

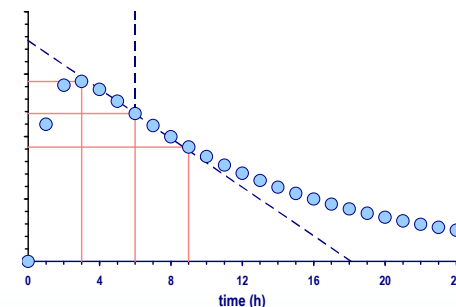
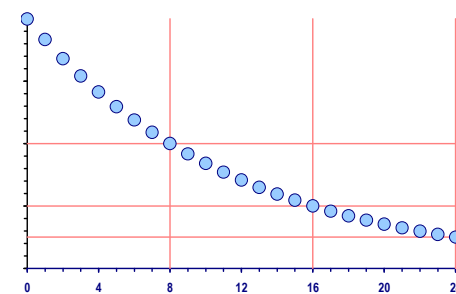
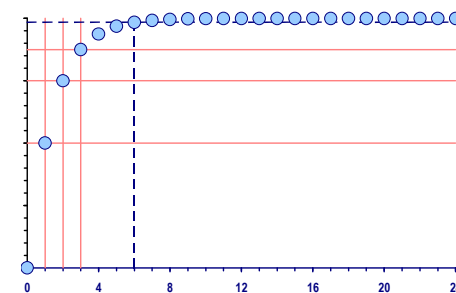
**Suggested if variability (especially in early time points) is high**

- **Multivariate statistical distance (MSD)**
  - MSD is estimated
    - Its 90% confidence interval calculated.
    - The upper limit compared to the similarity limit.
  - A subset of MSD is the Mahalanobis' Distance (MD).
    - Currently explored by the EMA's Biostatistical Working Party.
- **Model-dependent approaches**
  - Select a suitable model (quadratic, logistic, probit, Hill, Weibull, ...).
  - Similarity region is specified based on the variability.
  - Calculate MSD and CI as above.

# A(D)ME

*In vivo* curve can be described by absorption (A) and elimination (metabolization + excretion)

- One-compartment model does not have D (distribution).
  - Example:  $t_{1/2a}$  1 h,  $t_{1/2e}$  8 h
    - After  $3 \times t_{1/2a}$  ( 3 h) 87.5% are absorbed.
    - After  $3 \times t_{1/2e}$  (24 h) 87.5% are eliminated.
    - In the *in vivo* curve the inflection point (where the curve changes from concave to convex) is seen at  $2 \times t_{max}$  (6 h).  
At this time absorption is essentially complete (98.44%) and the *in vivo* curve practically represents elimination only.
- We can get *in vivo* absorption by subtracting the estimated elimination.



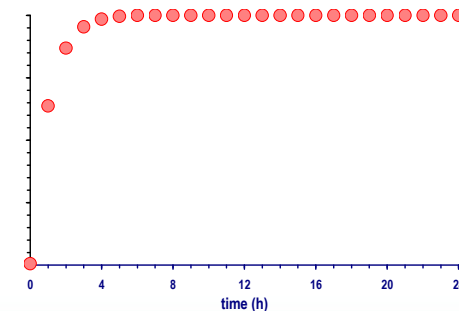
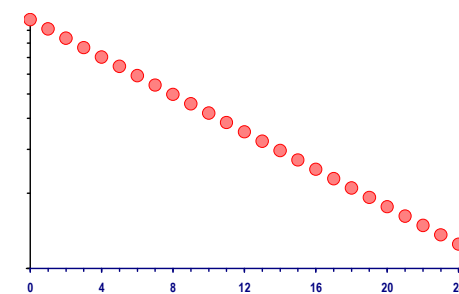
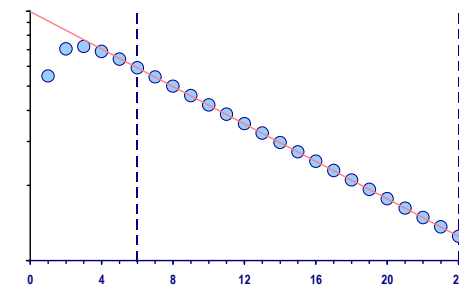
# A(D)ME

## Reconstructing *in vivo* absorption (residual method)

- Fit elimination ( $\lambda_z$  from  $2 \times t_{max}$  or later to  $t_z$ ).
- Predict *in vivo* elimination.
- *In vivo* absorption is the *in vivo* curve minus the predicted elimination.

## Different other methods exist.

- One-compartment model
  - Wagner-Nelson
$$abs(\%) = 100 \frac{C_t + k_{el} \cdot AUC_{0-t}}{k_{el} \cdot AUC_{0-\infty}}$$
- Two-compartment model
  - Loo-Riegelman (needs true elimination from iv); the distribution phase is reconstructed.

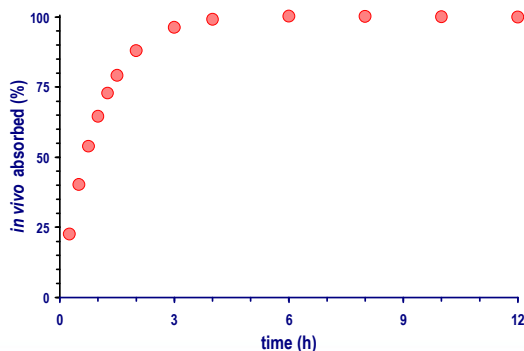
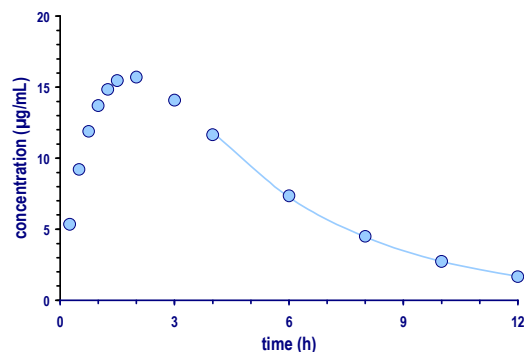




# Wagner-Nelson

$D$  100 mg,  $V$  4 L,  $F$  1,  $k_a$  1 h<sup>-1</sup> ( $t_{1/2}$  0.69 h),  $k_{el}$  0.25 h<sup>-1</sup> ( $t_{1/2}$  2.77 h)

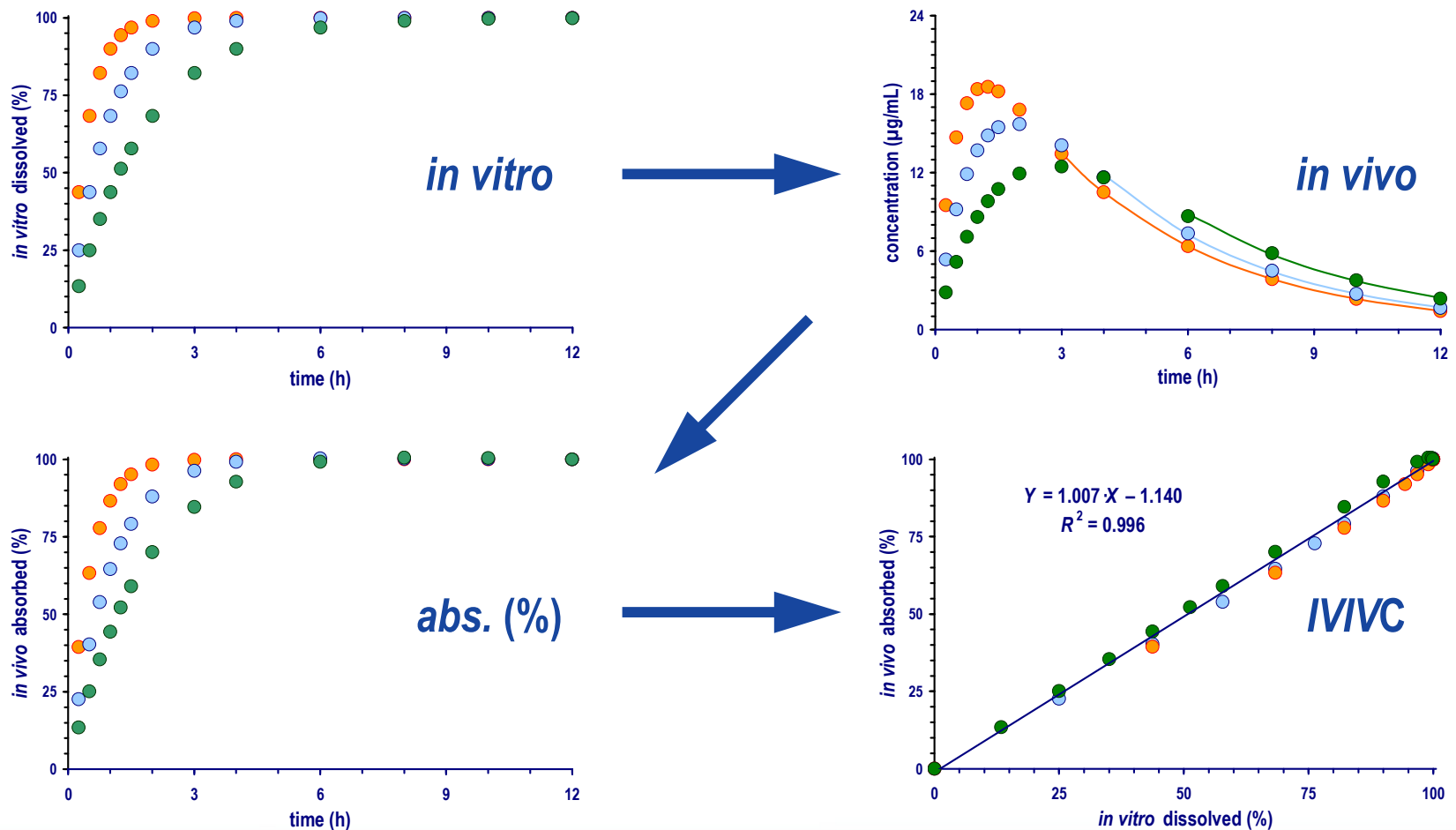
- Lin-up/log-down trapezoidal method for  $AUC_{0-t}$
- $\lambda_z$  (estimated from 4 to 12 hours) = 0.2444.
- $AUC_{0-\infty} = AUC_{0-12} + C_{12} / \lambda_z = 99.68$ .



$t$ (h)	$C$ (mg/mL)	$AUC_{0-t}$	abs (%)
0.00	BQL	–	–
0.25	5.35	0.67	22.63
0.50	9.20	2.49	40.26
0.75	11.89	5.12	53.94
1.00	13.70	8.32	64.58
1.25	14.84	11.89	72.84
1.50	15.47	15.68	79.22
2.00	15.71	23.47	88.03
3.00	14.09	38.36	96.31
4.00	11.65	51.19	99.17
6.00	7.36	69.87	100.31
8.00	4.50	81.50	100.23
10.00	2.73	88.88	100.08
12.00	1.66	92.68	100.00

# IVIVC (Level A)

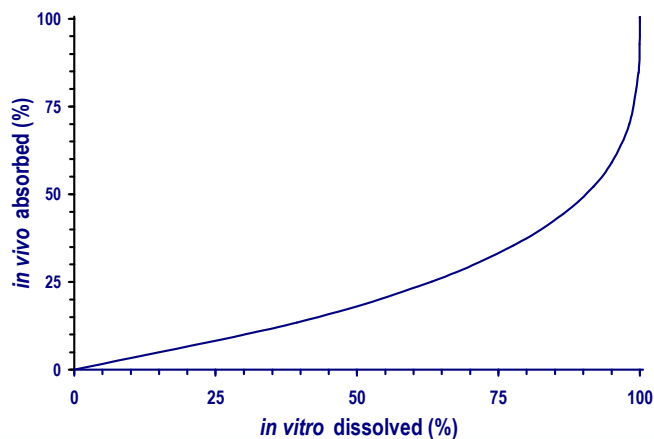
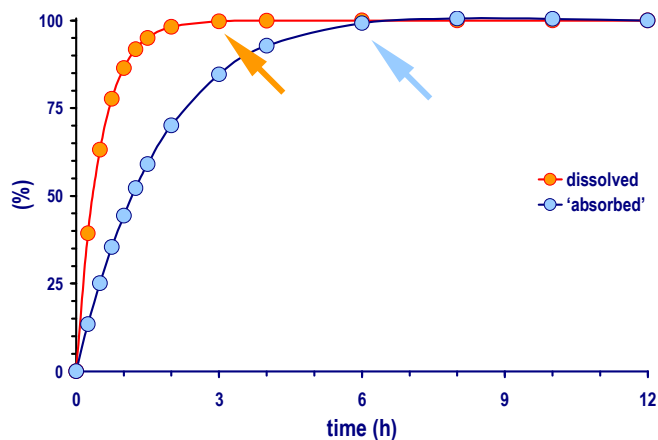
Three candidate formulations (**fast**, **intermediate**, **slow**)



# IVIVC (Level A)

## Different rates *in vitro* / *in vivo*

- Not suitable for IVIVC (nonlinear relationship)



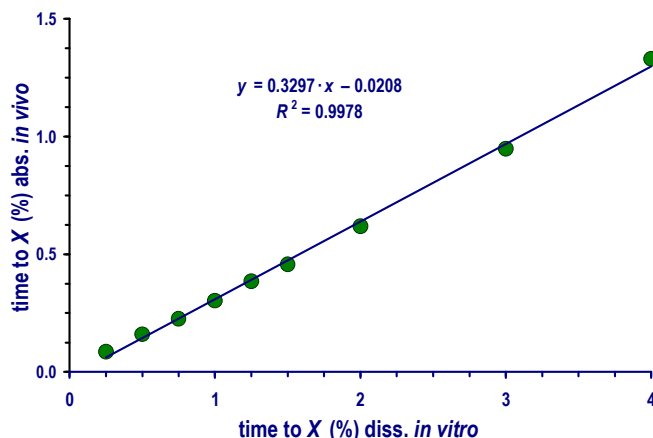
<i>t</i> (h)	<i>diss</i> (%)	<i>abs</i> (%)
0.00	0.00	0.00
0.25	39.35	13.44
0.50	63.21	25.14
0.75	77.69	35.44
1.00	86.47	44.37
1.25	91.79	52.22
1.50	95.02	59.04
2.00	98.17	70.10
3.00	99.75	84.66
4.00	99.97	92.82
6.00	100.00	99.27
8.00	100.00	100.57
10.00	100.00	100.43
12.00	100.00	100.00



# IVIVC (Level A)

## Different rates *in vitro* / *in vivo*

- Modify the dissolution method (e.g., less agitation) to get a better match.
- Establish a Levy plot (time to get % dissolved or absorbed). Use interpolation to find dissolution times which match absorption.



<i>in vivo</i>		<i>diss. time</i>	
<i>t</i> (h)	<i>abs</i> (%)	(h)	(h:mm)
0.00	0.00	0.00	0:00
0.25	13.44	0.06	0:03
0.50	25.14	0.14	0:08
0.75	35.44	0.23	0:13
1.00	44.37	0.31	0:18
1.25	52.22	0.39	0:23
1.50	59.04	0.47	0:28
2.00	70.10	0.64	0:38
3.00	84.66	0.97	0:58
4.00	92.82	1.30	1:17
6.00	99.27	1.96	1:57

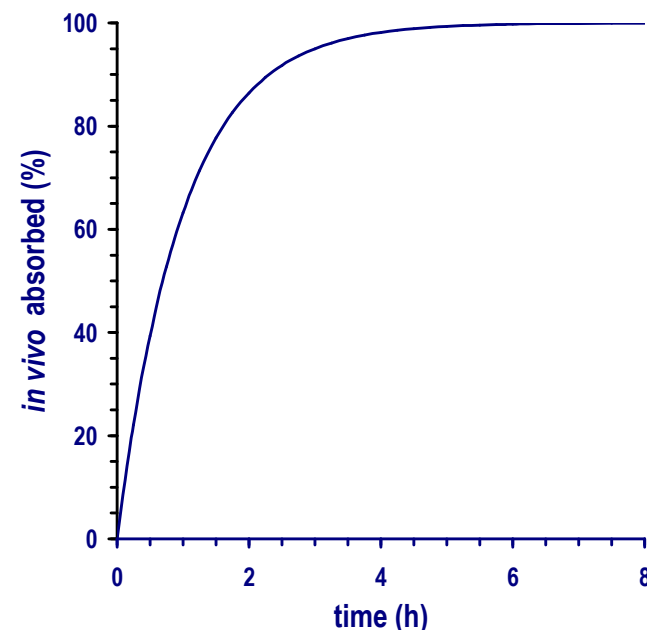
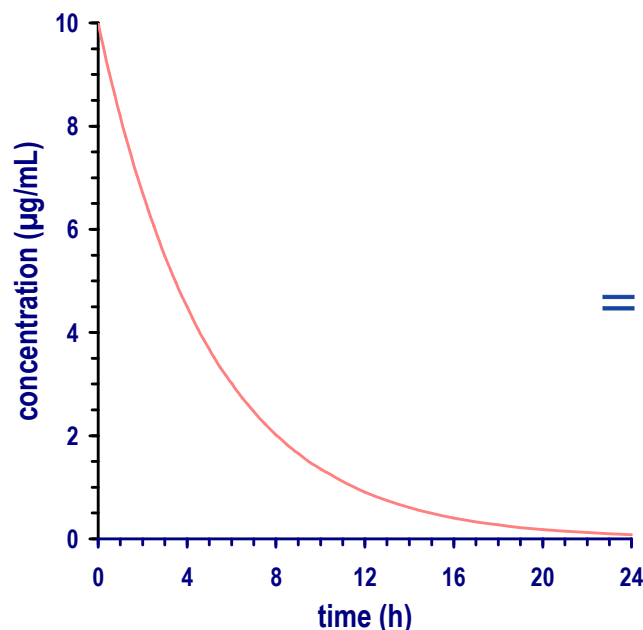
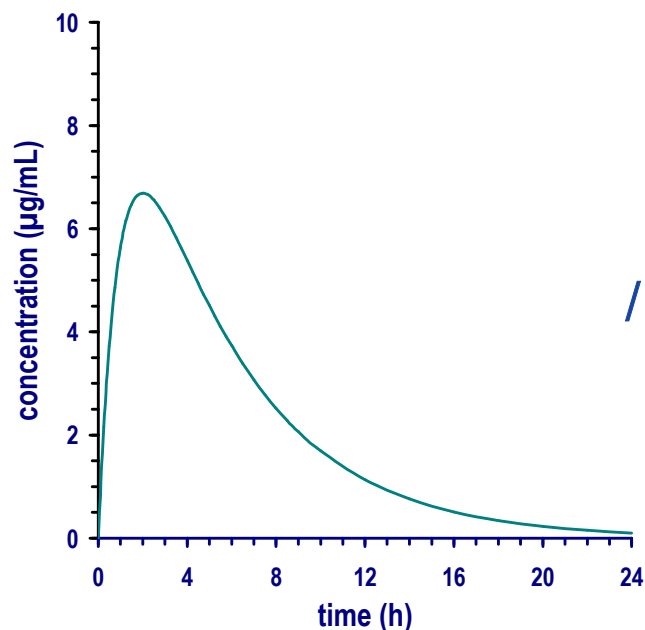
- Calculate new *in vitro* sampling times.

$$t_{in\ vitro} = t_{in\ vivo} \times 0.3297 - 0.0208.$$

# IVIVC (Level A)

## Alternative to Wagner-Nelson and Loo-Riegelman

- Deconvolution: Derive *in vivo* input curve from *in vivo* profile. Only method which is can be applied if there are more than two compartments. Notation:  $f = g / h$

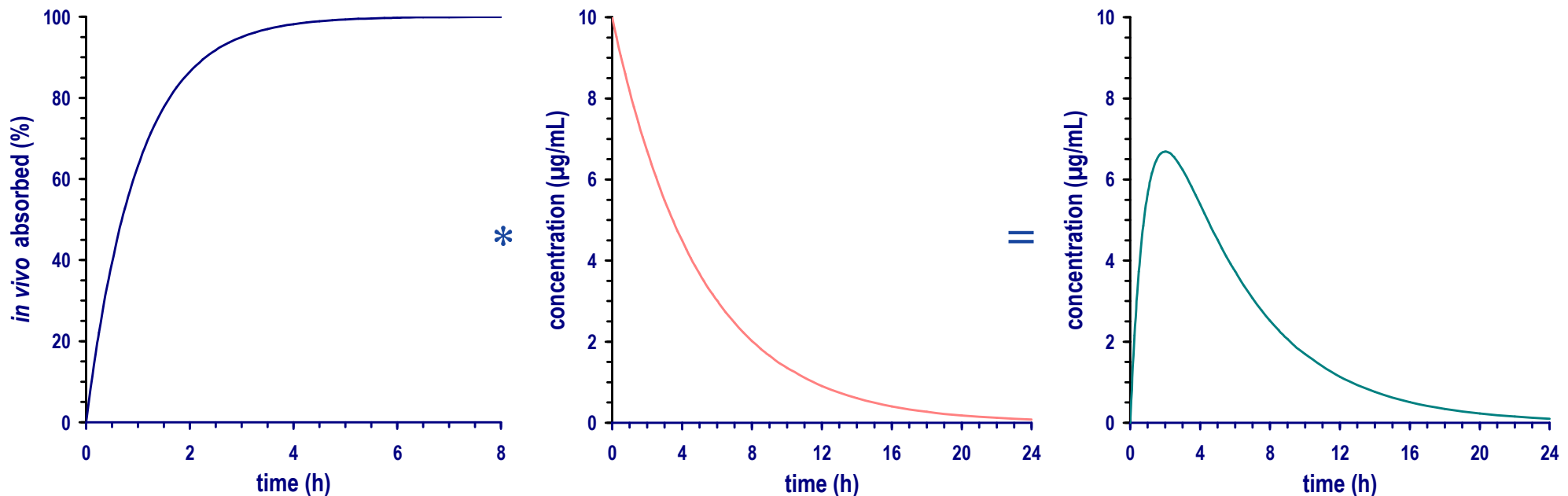


Jean-Michel Cardot. IVIVC Workshop. Mumbai, 27 – 29 January 2012.

# IVIVC (Level A)

## Alternative to Wagner-Nelson and Loo-Riegelman

- Convolution:** Derive *in vivo* profile from simulated *in vivo* input curve (obtained by IVIVC).  
 Notation:  $f = g * h$



Jean-Michel Cardot. *IVIVC Workshop*. Mumbai, 27 – 29 January 2012.

# IVIVC (Level A)

## Deconvolution / Convolution

- Already mathematically demanding for continuous functions – even more complicated if only data-pairs are available.
  - Numeric methods require equidistant supporting points. Must interpolate / impute data.
  - Requires additionally to % absorbed the rate of absorption  $dA / dt$  (method by Vaughan, Denis 1978).
  - Requires between six and ten sampling points in the absorption phase ( $\leq 2 \times t_{max}$ ).

# IVIVC (Levels B and C)

## Level B

- Correlation of statistical moments describing *in vitro* and *in vivo* profiles.
  - Mean dissolution time (*MDT*) with mean residence time and mean absorption time (*MRT*, *MAT*).  
Problem: *MRT* of *in vivo* profiles depend to a large part on distribution / elimination. Needs iv (or at least solution) data to obtain *MAT*.

## Level C

- Correlation of single-point metrics.
  - % dissolved (at least 80%) up to an certain time point with a PK metrics (e.g.,  $C_{max}$ , truncated *AUC*).
  - Few 'working' examples (e.g., glibenclamide).



# IVIVC

Quite often what one thinks to be ‘different’ (based on a QC dissolution method) turns out to be similar *in vivo*.

- Modify formulations, perform *in vivo* pilot studies until you see a difference *there*.
  - Then (!) develop a discriminatory *in vitro* method (Session 8) which is able to predict *in vivo* absorption
    - Try different agitation speeds, use surfactants, change the apparatus, if nothing helps – explore biorelevant media.
    - The final *in vitro* method likely has nothing in common with the one used in QC.  
*If Earl Grey with a sip of milk is predictive, use it!* (Jean-Michel Cardot)
- Once you found a discriminatory method, modify formulations to find one which matches the reference.
  - This does not guarantee that the reference will behaves *in vivo* like your best candidate.  
Another pilot (T vs. R) makes sense (to estimate *CV* and *GMR*).

# Similarity, Comparability and Correlation

**Thank You!**  
*Open Questions?*



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