



### $\alpha$ and $\beta$

- All formal decisions are subjected to two types of error:
  - $\blacksquare \alpha$  Probability of Error Type I (aka Risk Type I)
  - $\beta$  Probability of Error Type II (aka Risk Type II) Example from the justice system:

Verdict	Defendant innocent	Defendant guilty
Presumption of innocence not accepted (guilty)	Error type I	Correct
Presumption of innocence accepted (not guilty)	Correct	Error type II



#### $\alpha$ and $\beta$

#### Or in more statistical terms:

Decision	Null hypothesis true	Null hypothesis false
Null hypothesis rejected	Error type I	Correct (H <sub>a</sub> )
Failed to reject null hypothesis	Correct (H <sub>0</sub> )	Error type II

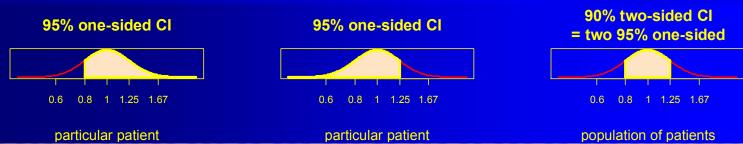
# •In BE-testing the null hypothesis is bioinequivalence $(\mu_1 \neq \mu_2)!$

Decision	Null hypothesis true	Null hypothesis false
Null hypothesis rejected	Patients' risk	Correct (BE)
Failed to reject null hypothesis	Correct (not BE)	Producer's risk



#### $\alpha$ ...

- •Patient's Risk to be treated with an inequivalent formulation ( $H_0$  falsely rejected)
  - BA of the test compared to reference in a *particular* patient is risky *either* below 80% *or* above 125%.
  - If we keep the risk of particular patients at  $\alpha$  0.05 (5%), the risk of the entire population of patients (<80% and >125%) is  $2 \times \alpha$  (10%) expressed as: 90% CI =  $1 2 \times \alpha = 0.90$





### $\dots$ and $\beta$

- Producer's Risk to get no approval for a equivalent formulation (H<sub>0</sub> falsely not rejected)
  - Set in study planning to  $\le 0.2$  (20%), where power =  $1 \beta = \ge 80\%$
  - If power is set to 80 %,
    one out of five studies will fail just by chance!

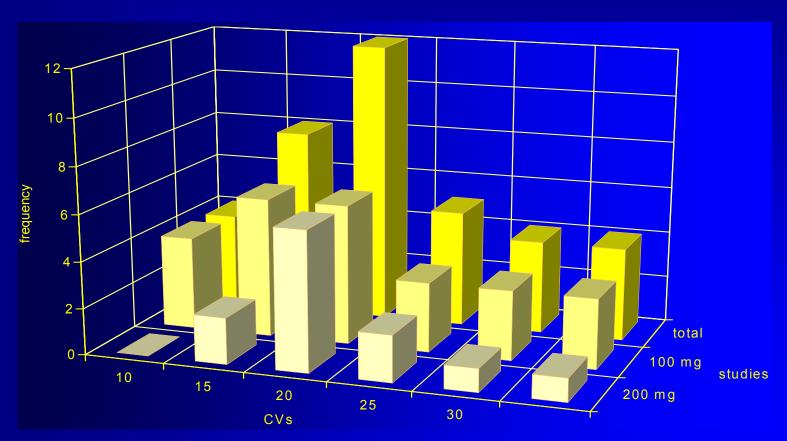
$$\alpha \, 0.05$$
BE

not BE
 $\beta \, 0.20$ 
 $0.20 = 1/5$ 

■ A posteriori (post hoc) power does not make sense! Either a study has demonstrated BE or not.



#### **Published data**



**Doxicycline** (37 studies from **Blume/Mutschler**, *Bioäquivalenz*: Qualitätsbewertung wirkstoffgleicher Fertigarzneimittel, GOVI-Verlag, Frankfurt am Main/Eschborn, 1989-1996)





#### Sample Size (Guidelines)

- Recommended minimum
  - ■12 WHO, EU, CAN, NZ, AUS, AR, MZ, ASEAN States, RSA, Russia (2011 Draft)
  - 12 USA 'A pilot study that documents BE can be appropriate, provided its design and execution are suitable and a sufficient number of subjects (*e.g.*, 12) have completed the study.'
  - ■18 Russia (2008)
  - 20 RSA (MR formulations)
  - 24 Saudia Arabia (12 to 24 if statistically justifiable)
  - 24 Brazil
  - 'Sufficient number' Japan





#### Sample Size (Limits)

#### Maximum

- NZ: If the calculated number of subjects appears to be higher than is ethically justifiable, it may be necessary to accept a statistical power which is less than desirable. Normally it is not practical to use more than about 40 subjects in a bioavailability study.
- All others: Not specified (judged by IEC/IRB or local) Authorities).
  - ICH E9, Section 3.5 applies: "The number of subjects in a clinical trial should always be large enough to provide a reliable answer to the questions addressed."





#### **EMEA**

- NfG on the Investigation of BA/BE (2001)
  - The number of subjects required is determined by
    - the error variance associated with the primary characteristic to be studied as estimated from
      - > a pilot experiment,
      - > previous studies, or
      - published data,
    - the significance level desired,
    - the expected deviation ( $\Delta$ ) from the reference product compatible with BE and,
    - the required power.





#### **EMA**

- BE Guideline (2010)
  - The number of subjects to be included in the study should be based on an

appropriate

sample size calculation.

Cookbook?



#### **Tools**

- Sample Size Tables (Phillips, Diletti, Hauschke, Chow, Julious, ...)
- Approximations (Diletti, Chow, Julious, ...)
- General purpose (SAS, S+, R, StaTable, ...)
- Specialized Software (nQuery Advisor, PASS, FARTSSIE, StudySize, ...)
- Exact method (Owen implemented in R-package PowerTOST)\*

<sup>\*</sup> Thanks to Detlew Labes!



### **Approximations obsolete**

- Exact sample size tables still useful in checking plausibility of software's results
- Approximations based on noncentral t (FARTSSIE17)



http://individual.utoronto.ca/ddubins/FARTSSIE17.xls

or  $\mathbb{R} / S+ \rightarrow$ 

Exact method (Owen) in R-package PowerTOST

```
http://cran.r-project.org/web/packages/PowerTOST/
require(PowerTOST)
    sampleN.TOST(alpha=0.05,
    targetpower=0.80, theta0=0.95,
    CV=0.30. design='2x2')
```

```
alpha
        <- 0.05
        <- 0.30
                     # intra-subject CV
CV
theta1 <- 0.80
                     # lower acceptance limit
theta2 <- 1/theta1 # upper acceptance limit
                    # expected ratio T/R
theta0 <- 0.95
                     # minimum power
PwrNeed <- 0.80
Limit
        <- 1000
                     # Upper Limit for Search
                    # start value of sample size search
        <- 4
        <- sqrt(2)*sqrt(log(CV^2+1))
repeat{
        <- qt(1-alpha,n-2)
        <- sqrt(n)*(log(theta0)-log(theta1))/s
  nc1
        <- sqrt(n)*(log(theta0)-log(theta2))/s
  prob1 \leftarrow pt(+t,n-2,nc1); prob2 \leftarrow pt(-t,n-2,nc2)
  power <- prob2-prob1
                    # increment sample size
  if(power >= PwrNeed | (n-2) >= Limit) break }
       <-n-2
if(Total == Limit){
  cat('Search stopped at Limit', Limit,
        obtained Power', power*100, '%\n')
  cat('Sample Size', Total, '(Power', power*100, '%)\n')
```



#### Which Power?

- Generally Producer's Risk 10–20%
  - Plan for 90% allowing for contingency e.g.,
    - drop-outs,
    - CV<sub>intra</sub> higher than assumed,
    - deviation of test from reference larger than expected.
  - Power >90% might lead to ethical problems ('forced bioequivalence').
  - ■FDA (2001): 80–90%
  - ■EMA (2010): 'appropriate'...
  - Russia (2008, 2011 draft): ≥80%



### **End of the Story?**

- 'Doing the maths' is just part of the job!
  - Does it make sense to rely on studies of different origin and sometimes unknown quality?
    - The reference product may have been subjected to many (minor only?) changes from the formulation used in early publications.
    - Different bioanalytical methods are applied. Newer (e.g. LC/MS-MS) methods are not necessarily better in terms of variability.
    - Generally insufficient information about the clinical setup (e.g., posture control).
    - Review studies critically; don't try to mix oil with water.





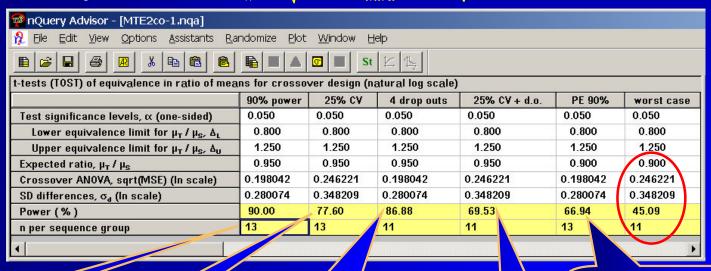
- •ICH E9 (1998)
  - Section 3.5 Sample Size, paragraph 3
    - The method by which the sample size is calculated should be given in the protocol [...]. The basis of these estimates should also be given.
    - It is important to investigate the sensitivity of the sample size estimate to a variety of deviations from these assumptions and this may be facilitated by providing a range of sample sizes appropriate for a reasonable range of deviations from assumptions.
    - In confirmatory trials, assumptions should normally be based on published data or on the results of earlier trials.





Example

nQuery Advisor:  $\sigma_w = \sqrt{\ln(CV_{intra}^2 + 1)}; \sqrt{\ln(0.2^2 + 1)} = 0.198042$ 



20% CV: n=26

25% CV: power 90% → **78%** 

20% CV, 4 drop outs: power  $90\% \rightarrow 87\%$ 

25% CV, 4 drop outs: power  $90\% \rightarrow 70\%$ 

20% CV, PE 90%: power 90%  $\rightarrow$  67%





#### Example

PowerTOST, function sampleN.TOST







 To estimate Power for a given sample size, use function *power*. TOST

```
require(PowerTOST)
power.TOST(theta0=0.95, CV=0.25, n=26)
[1] 0.7760553
power.TOST(theta0=0.95, CV=0.20, n=22)
[1] 0.8688866
power.TOST(theta0=0.95, CV=0.25, n=22)
[1] 0.6953401
power.TOST(theta0=0.90, CV=0.20, n=26)
[1] 0.6694514
power.TOST(theta0=0.90, CV=0.25, n=22)
[1] 0.4509864
```

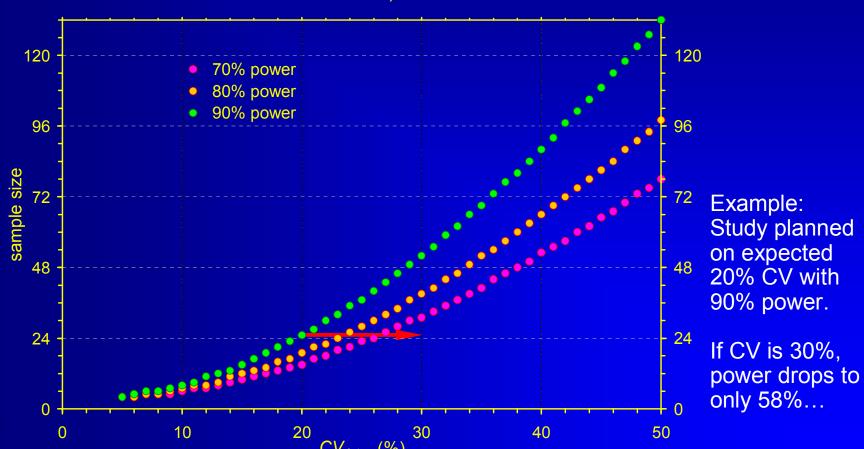






### CV based on assumptions!

2×2 cross-over, T/R 0.95





## Add-on / Two-Stage Designs

- Sometimes properly designed and executed studies fail due to
  - "true' bioinequivalence,
  - poor study conduct (increasing variability),
  - pure chance (producer's risk hit),
  - false (over-optimistic) assumptions about variability and/or T/R-ratio.
- The patient's risk must be preserved
  - Already noticed at Bio-International Conferences (1989, 1992) and guidelines from the 1990s.



### **Sequential Designs**

- Have a long and accepted tradition in clinical research (mainly phase III)
  - Based on work by Armitage *et al.* (1969), McPherson (1974), Pocock (1977), O'Brien and Fleming (1979), Lan & DeMets (1983), ...
    - First proposal by Gould (1995) in the area of BE did not get regulatory acceptance in Europe, but
    - new methods stated in recent guidelines.

#### **AL Gould**

Group Sequential Extension of a Standard Bioequivalence Testing Procedure J Pharmacokin Biopharm 23/1, 57–86 (1995)



### **Sequential Designs**

- Methods by Potvin et al. (2008) promising
  - Supported by the 'Product Quality Research Institute' (members: FDA/CDER, Health Canada, USP, AAPS, PhRMA...)
    - Three of BEBAC's protocols accepted by German BfArM, one product approved in 06/2011.

Potvin D, Diliberti CE, Hauck WW, Parr AF, Schuirmann DJ, and RA Smith Sequential design approaches for bioequivalence studies with crossover designs Pharmaceut Statist 7/4, 245–62 (2008), DOI: 10.1002/pst.294
http://www3.interscience.wiley.com/cgi-bin/abstract/115805765/ABSTRACT



#### **Review of Guidelines**

- Canada (May 2012)
   Potvin et al. Method C recommended.
- FDA (Jun 2012)
   Potvin et al. Method C recommended.
   API specific guidances: Loteprednol, Dexamethasone / Tobramycin.
- EMA (Jan 2010)
   Acceptable; Potvin et al. Method B preferred.
- Russia (Draft 2011)
   Acceptable (Methods B and C).



## Two-Stage Design

- •EMA GL on BE (2010)
  - Section 4.1.8
    - Initial group of subjects treated and data analysed.
    - If BE not been demonstrated an additional group can be recruited and the results from both groups combined in a final analysis.
    - Appropriate steps to preserve the overall type I error (patient's risk).
    - Stopping criteria should be defined a priori.
    - First stage data should be treated as an interim analysis.



## Two-Stage Design

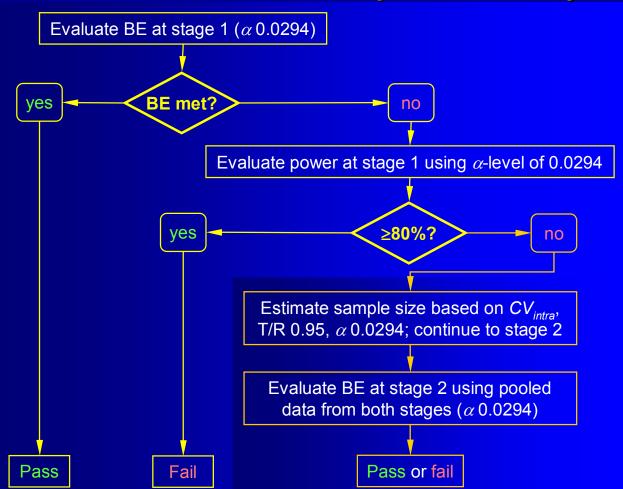
- •EMA GL on BE (2010)
  - Section 4.1.8 (cont'd)
    - ■Both analyses conducted at adjusted significance levels (with the confidence intervals accordingly using an adjusted coverage probability which will be higher than 90%). [...] 94.12% confidence intervals for both the analysis of stage 1 and the combined data from stage 1 and stage 2 would be acceptable, but there are many acceptable alternatives and the choice of how much alpha to spend at the interim analysis is at the company's discretion.



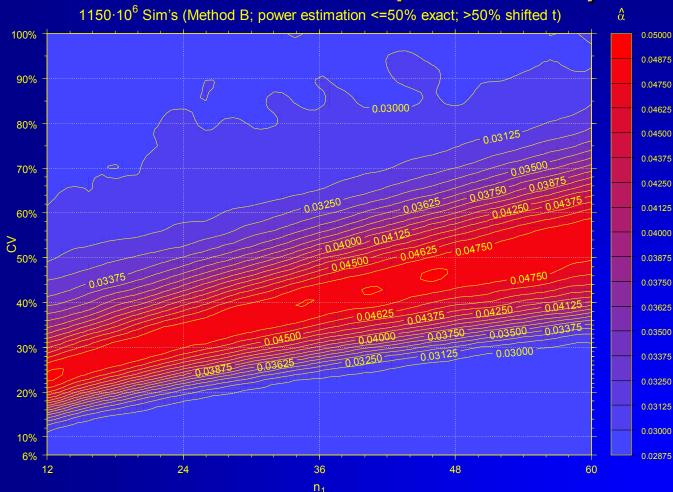
## Two-Stage Design

- •EMA GL on BE (2010)
  - Section 4.1.8 (cont'd)
    - Plan to use a two-stage approach must be prespecified in the protocol along with the adjusted significance levels to be used for each of the analyses.
    - When analysing the combined data from the two stages, a term for stage should be included in the ANOVA model.

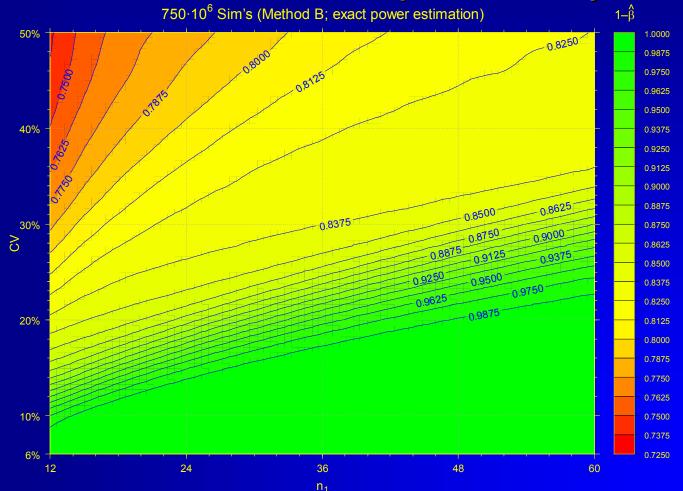






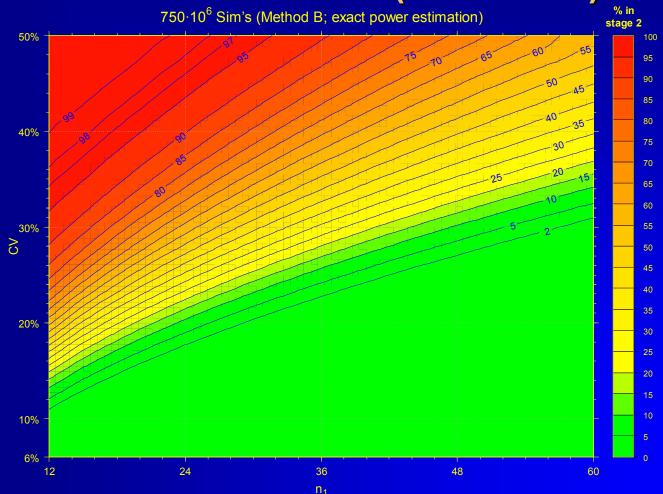




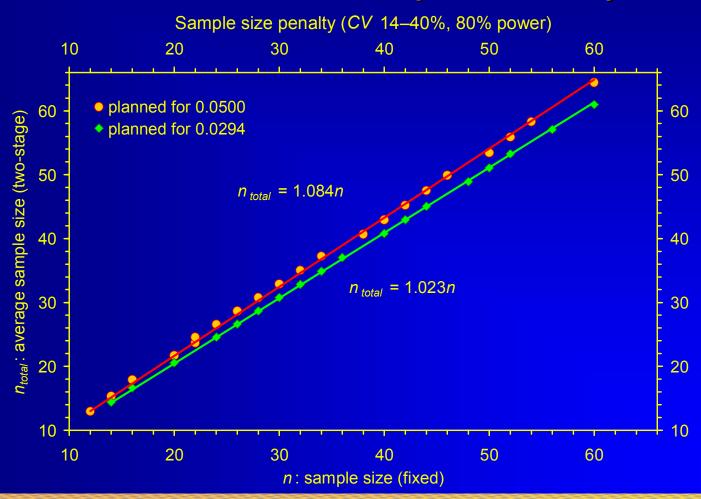














- Technical Aspects
  - Only one Interim Analysis (after stage 1).
  - Use software (wide step sizes in Diletti's tables); preferrable the exact method (avoid approximations).
  - Should be termed 'Interim Power Analysis' *not* 'Bioequivalence Assessment' in the protocol.
  - No a posteriori Power only a validated method in the decision tree.
  - No adjustment for T/R observed in stage 1 (not fully adaptive).



- Technical Aspects (cont'd)
  - No futility rule preventing to go into stage 2 with a very high sample size! Must be clearly stated in the protocol (unfamiliar to the IEC because common in Phase III).
  - Pocock's  $\alpha$  0.0294 is used in stage 1 and in the pooled analysis (data from stages 1 + 2), i.e., the 1 2× $\alpha$  = 94.12% CI is calculated.
  - Overall patient's risk preserved at ≤0.05.



- Technical Aspects (cont'd)
  - If the study is stopped after stage 1, the (conventional) statistical model is:

```
fixed: sequence + period + treatment
random: subject(sequence)
```

If the study continues to stage 2, the model for the combined analysis is:

```
fixed: sequence + stage + period(stage) + treatment
random: subject(sequence × stage)
```

No poolability criterion!
 Combining is always allowed – even if a significant difference between stages is observed. No need to test this effect.



- Technical Aspects (cont'd)
  - Potvin *et al.* used a simple approximative power estimation based on the shifted *t*-distribution.
  - If possible use the exact method (Owen; R package PowerTOST method = 'exact') or at least one based on the noncentral t-distribution (PowerTOST method = 'noncentral').
  - Power obtained in stage 1 (example 2 from Potvin):

method	power
approx. (shifted t)	50.49%
approx. (noncentral t)	52.16%
exact	52.51%



```
Model Specification and User Settings
                                                             12 subjects in stage 1,
      Dependent variable: Response
                                                             conventional BE model
                Transform: LN
              Fixed terms : int+Sequence+Period+Treatment
   Random/repeated terms : Sequence*Subject
Final variance parameter estimates:
   Var(Sequence*Subject)
                              0.408682
                                                CV<sub>intra</sub> 18.2%
           Var(Residual)
                              0.0326336
          Intrasubject CV
                             0.182132
Bioequivalence Statistics
                                                                    \alpha 0.0294
User-Specified Confidence Level for CI's = 94.1200
Percent of Reference to Detect for 2-1 Tests = 20.0%
A.H.Lower = 0.800 A.H.Upper = 1.250
Reference: Reference LSMean = 0.954668
                                         SE = 0.191772
                                                         GeoLSM = 2.597808
                       LSMean = 1.038626 SE = 0.191772 GeoLSM = 2.825331
Test:
          Test
                   0.0840, Diff_SE = 0.0737, df = 10.0
   Difference =
   Ratio(\%Ref) = 108.7583
                                             Failed with 94.12% Confidence Interval
                      Classical
                 92.9330, 127.2838)
   CI User = (
   Failed to show average bioequivalence for confidence=94.12 and percent=20.0.
```

38



# Potvin et al. (Method B)

```
α 0.0294, T/R 95% – not 108.76%
require(PowerTOST)
                                                 observed in stage 1!
power.TOST(alpha=0.0294, theta0=0.95,
                                                 CV_{intra} 18.2%, 12 subjects in stage 1
           CV=0.182132, n=12, design='2x2',
           method='exact')
                           Power 52.5% – initiate stage 2
[1] 0.5251476
sampleN.TOST(alpha=0.0294, targetpower=0.80, logscale=TRUE,
            theta1=0.8, theta2=1.25, theta0=0.95,
            CV=0.182132, design='2x2', method='exact',
            print=TRUE)
                                                    Estimate total sample size:
++++++++ Equivalence test - TOST ++++++++
            Sample size estimation
                                                    \alpha 0.0294, T/R 95%, CV_{intra} 18.2%,
                                                    80% power
Study design: 2x2 crossover
log-transformed data (multiplicative model)
alpha = 0.0294, target power = 0.8
                  = 0.8 ... 1.25
BE margins
Null (true) ratio = 0.95, CV = 0.182132
Sample size
       power
                           Total sample size 20: include another 8 in stage 2
20
     0.829160
```

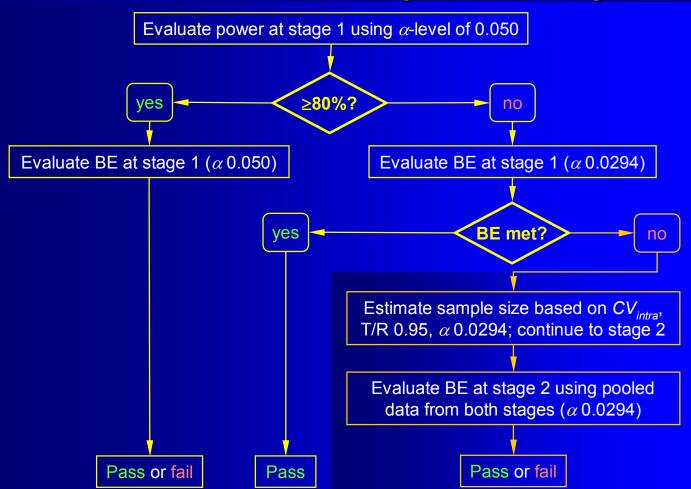


# Potvin et al. (Method B)

```
8 subjects in stage 2 (20 total),
Model Specification and User Settings
                                                  modified model in pooled analysis
      Dependent variable : Cmax (ng/mL)
                Transform: LN
              Fixed terms : int+Sequence+Stage+Period(Stage)+Treatment
   Random/repeated terms : Sequence*Stage*Subject
Final variance parameter estimates:
Var(Sequence*Stage*Subject)
                              0.518978
           Var(Residual)
                              0.0458956
          Intrasubject CV
                              0.216714
                                                                    \alpha 0.0294 in
Bioequivalence Statistics
                                                                    pooled analysis
User-Specified Confidence Level for CI's = 94.1200
Percent of Reference to Detect for 2-1 Tests = 20.0%
A.H.Lower = 0.800 A.H.Upper = 1.250
Formulation variable: Treatment
Reference: Reference LSMean = 1.133431 SE = 0.171385 GeoLSM = 3.106297
                       LSMean = 1.147870 SE = 0.171385 GeoLSM = 3.151473
Test:
          Test
                   0.0144, Diff_SE = 0.0677, df = 17.0
   Difference =
   Ratio(\%Ref) = 101.4544
                                                       BE shown with 94.12% CI:
                      Classical
                                                       overall \alpha \leq 0.05!
   CI 90\% = (
                 90.1729, 114.1472)
                 88.4422, 116.3810)
   CI User = (
    Average bioequivalence shown for confidence=94.12 and percent=20.0.
```

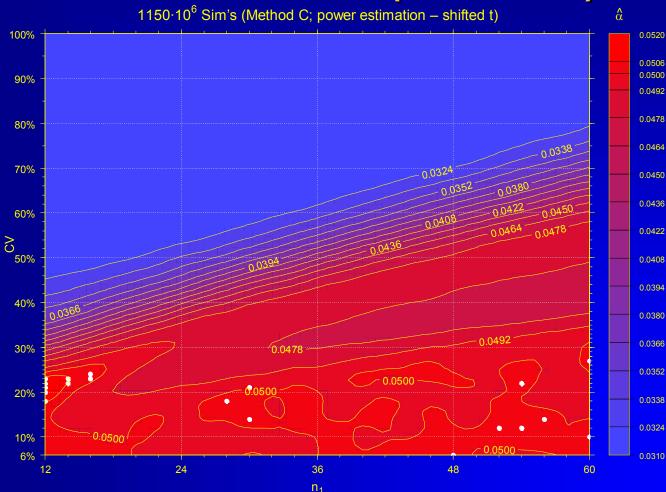


# Potvin et al. (Method C)





# Potvin et al. (Method C)





# Potvin et al. (Method B vs. C)

- Pros & cons
  - ■Method C (*if power*  $\geq$ 80%!) is a conventional BE study; no penality in terms of  $\alpha$  needs to be applied.
  - Method C proceeds to stage 2 less often and has smaller average total sample sizes than Method B for cases where the initial sample size is reasonable for the CV.
  - If the size of stage 1 is low for the actual *CV* both methods go to stage 2 almost all the time; total sizes are similar.
  - Method B slightly more conservative than C.



# Potvin et al. (Method B vs. C)

- Recommendations
  - Method C preferred due to slightly higher power than method B (FDA, HPB). Method B for EMA.
  - Plan the study as if the CV is known
    - If assumptions turn out to be true = no penalty
    - If lower power ( $CV_{intra}$  higher than expected), BE still possible in first stage (penalty; 94.12% CI) or continue to stage 2 as a 'safety net'.
  - ■Don't jeopardize! Smaller sample sizes in the first stage than in a fixed design don't pay off.

    Total sample sizes are ~10–20% higher.



# Sequential Designs

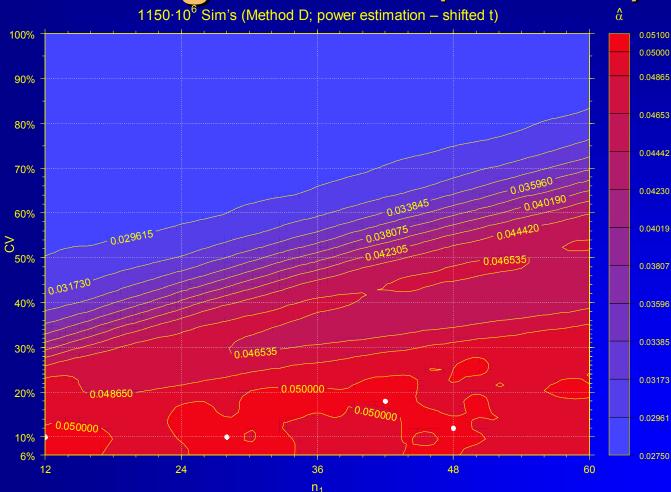
- Methods by Potvin et al. (2008) limited to T/R of 0.95 and 80% power
  - Follow-up paper 2011
    - T/R 0.90 instead of 0.95.
    - Method D (like C, but  $\alpha$  0.0280 instead of  $\alpha$  0.0294).
    - Might be useful if T/R 0.95 and power 90% as well; not validated yet! Simulations required.

Montague TH, Potvin D, DiLiberti CE, Hauck WW, Parr AF, and DJ Schuirmann Additional results for 'Sequential design approaches for bioequivalence studies with crossover designs'

Pharmaceut Statist 1/1, 8–13 (2011), DOI: 10.1002/pst.483



# Montague et al. (Method D)





# Case Studies (EMA)

- Method C: Study passed in first stage (49 subjects, CV 30.65%, 90% CI)
  - Deficiency 1: Unadjusted  $\alpha$  in stage 1 not acceptable
    - Response 1: Study passed with 94.12% CI (post hoc switch to Method B).
  - Deficiency 2: The Applicant should demonstrate that the type I error inflation which can be expected from the chosen approach, did not impact on the decision of bioequivalence.
    - Response 2: One million simulations based on study's sample size and CV.  $\alpha_{emp}$  0.0494 (95% CI: 0.0490 0.0498)



# Case Studies (EMA)

- Method C: Study stopped in first stage
   AUC power >80%, passed with 90% CI
   C<sub>max</sub> power <80%, passed with 94.12% CI</li>
  - Deficiency: Adapting the confidence intervals based upon power is not acceptable and also not in accordance with the EMA guideline. Confidence intervals should be selected a priori, without evaluation of the power. Therefore, the applicant should submit the 94.12% confidence intervals for AUC.
    - Pending: AUC fails with 94.12% CI...

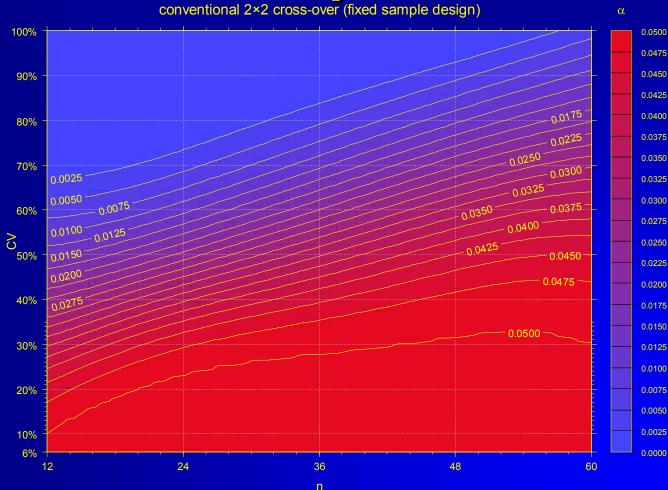


## Outlook

- Feasibility / futility rules.
- Arbitrary expected T/R and/or power.
- Methods without interim power.
- Application to parallel designs.
- Dropping a candidate formulation from a higher-order cross-over; continue with 2×2.
- Exact method (not requiring simulations).
- Adaption for T/R observed in stage 1 (full adaptive design).



# Don't panic!





# Thank You! Two-Stage Designs Open Questions?



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Dedicated to the memory of Dirk Maarten Barends (1945 – 2012).



### To bear in Remembrance...

Power. That which statisticians are always calculating but never have.

Power Calculation – A guess masquerading as mathematics. Stephen Senn





In bioequivalence we must not forget the only important – *the patient*! He/she is living person, not just  $\alpha$  0.05.

Dirk Marteen Barends

It is a good morning exercise for a research scientist to discard a pet hypothesis every day before breakfast.

It keeps him young.

Konrad Lorenz







#### References

#### •ICH

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- EMA-CPMP/CHMP/EWP
  - Points to Consider on Multiplicity Issues in Clinical Trials (2002)
  - Guideline on the Investigation of BE (2010)
  - Questions & Answers: Positions on specific questions addressed to the EWP therapeutic subgroup on Pharmacokinetics (2011, 2012)

#### •US-FDA

- Center for Drug Evaluation and Research (CDER)
  - Statistical Approaches Establishing Bioequivalence (2001)
  - Bioequivalence Recommendations for Specific Products (2007-2012): Draft Guidance on Lotepredenol (Jun 2012) Draft Guidance on Dexamethasone/Tobramycin (Jun 2012)
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- AL Gould Group Sequential Extension of a Standard Bioequivalence Testing Procedure J Pharmacokin Biopharm 23/1, 57-86 (1995) DOI: 10.1007/BF02353786



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- García-Arieta A and J Gordon Bioequivalence Requirements in the European Union: Critical Discussion
- The AAPS Journal 14/4, 738-48 (2012) ■ BM Davit Sequential Designs and Interim Analyses in Bioequivalence: FDA's Experience Mini-Symposium on Adaptive Study Designs and Assessment Approaches for Bioequivalence AAPS Annual Meeting, Chicago, IL, October 13-18, 2012

